

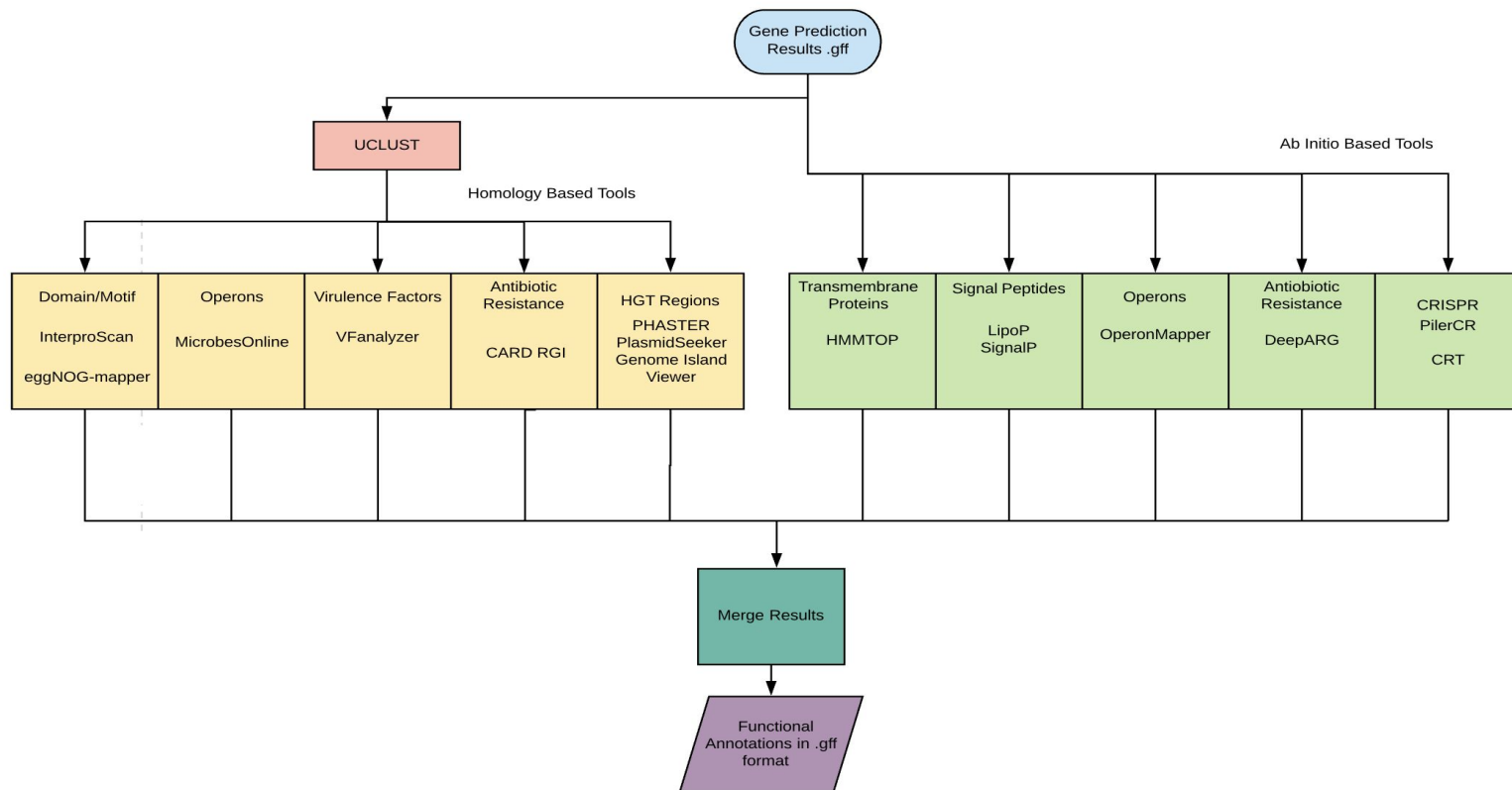
# Functional Annotation Results

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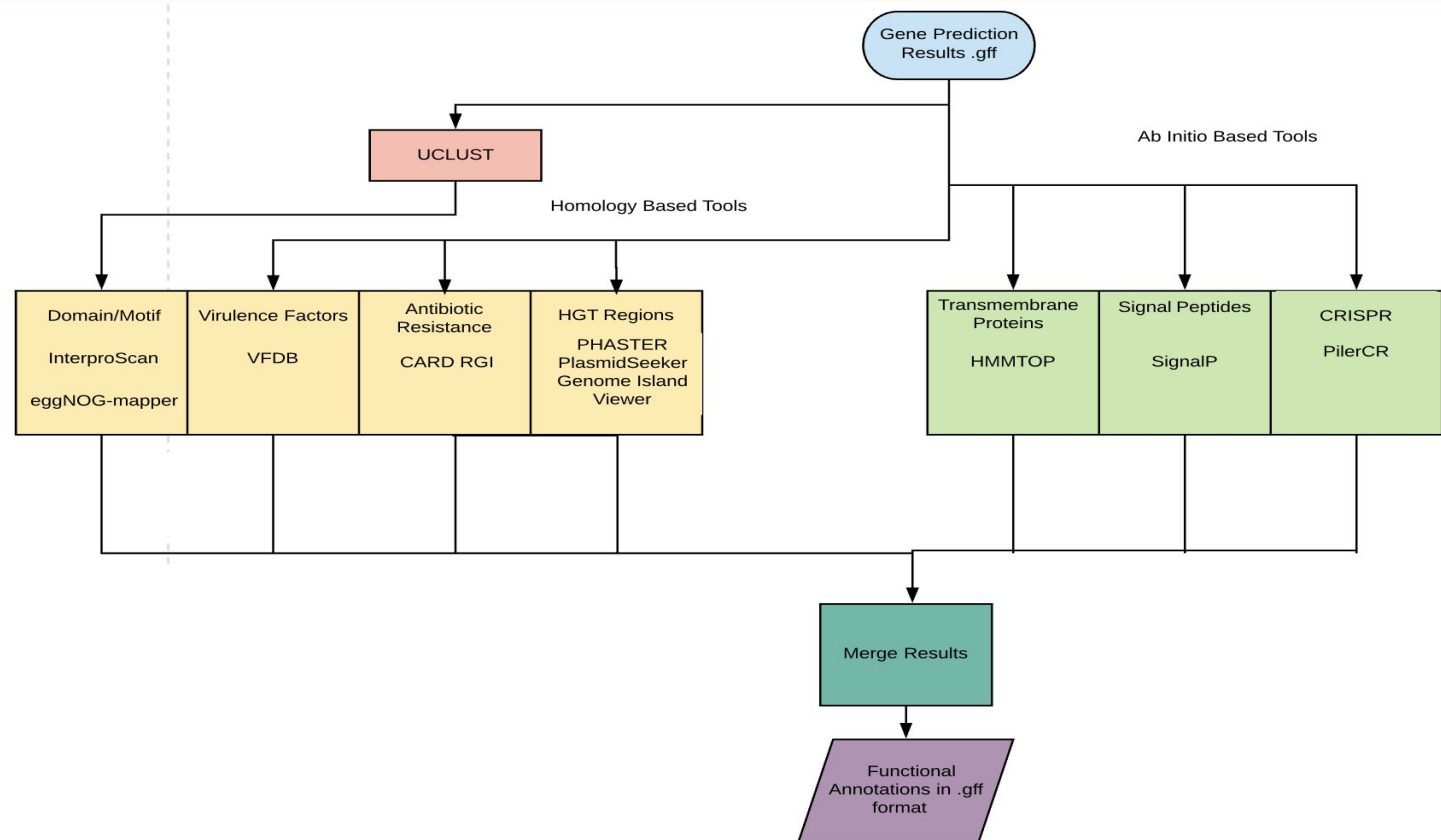
Team 3

Allison Rozanski, Gulay Bengu Ulukaya, Cheng Shen-Yi,  
Pallavi Misra

# Initial Pipeline



# Updated Pipeline



# UCLUST<sub>[2]</sub>

Each cluster contains a single centroid sequence upon which the other sequences must have a certain sequence similarity to to be considered apart of the cluster [2]

The threshold limit can be thought of as the radius of the cluster

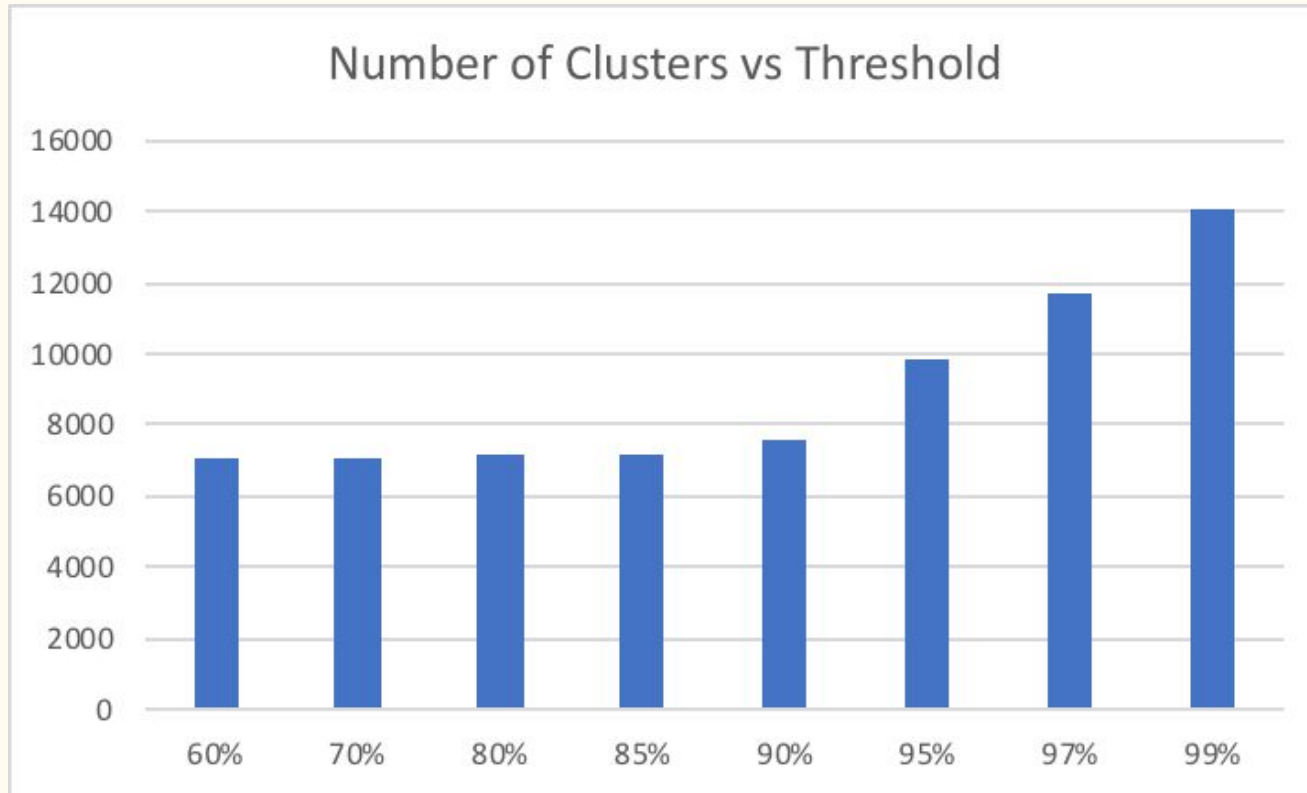
Identities are determined by global alignment

Used a 97% identity cluster threshold: obtains larger average cluster sizes and relatively low amount of singletons: 1%

Although 95% identity had larger average cluster sizes we wanted a higher degree of specificity in our results.

Identity Threshold	Clusters	Singletons	Avg Cluster Size
99%	14,010	976	11
97%	11,720	459	14.2
95%	9,871	322	15.6

# UCLUST: Cluster Frequencies



# eggNOG Mapper<sub>[1]</sub>

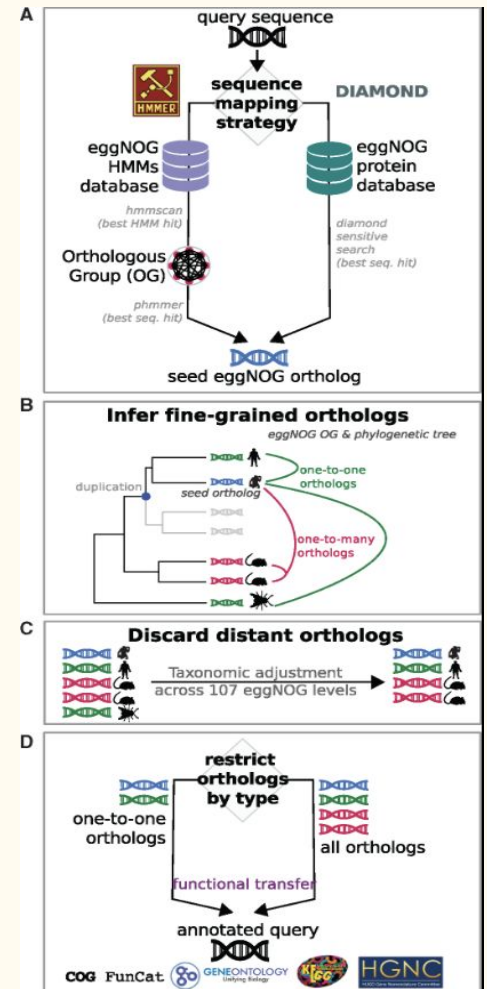
Download eggNOG databases:

```
download_eggnog_data.py bact
```

Run eggNOG:

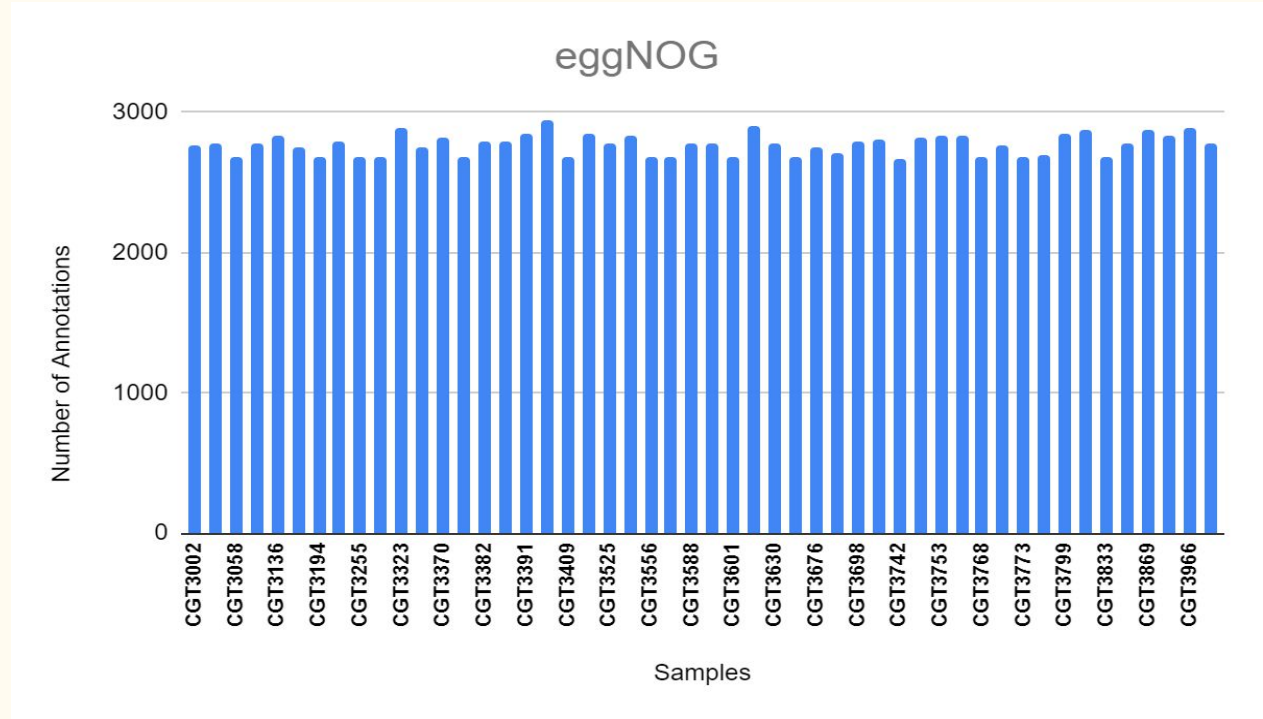
- DIAMOND vs. HMMER
- “recommended for very large data sets such as metagenomes, as well as for annotating organisms with close relatives among the species covered by eggNOG”
- Because the eggNOG database covers close relatives of *Listeria monocytogenes* and due to the size of our analysis even after clustering we decided to utilize DIAMOND

```
emapper.py -i ../USEARCH/All_Centroid.fasta -m diamond  
--translate -d bact -o eggNog_annotations
```



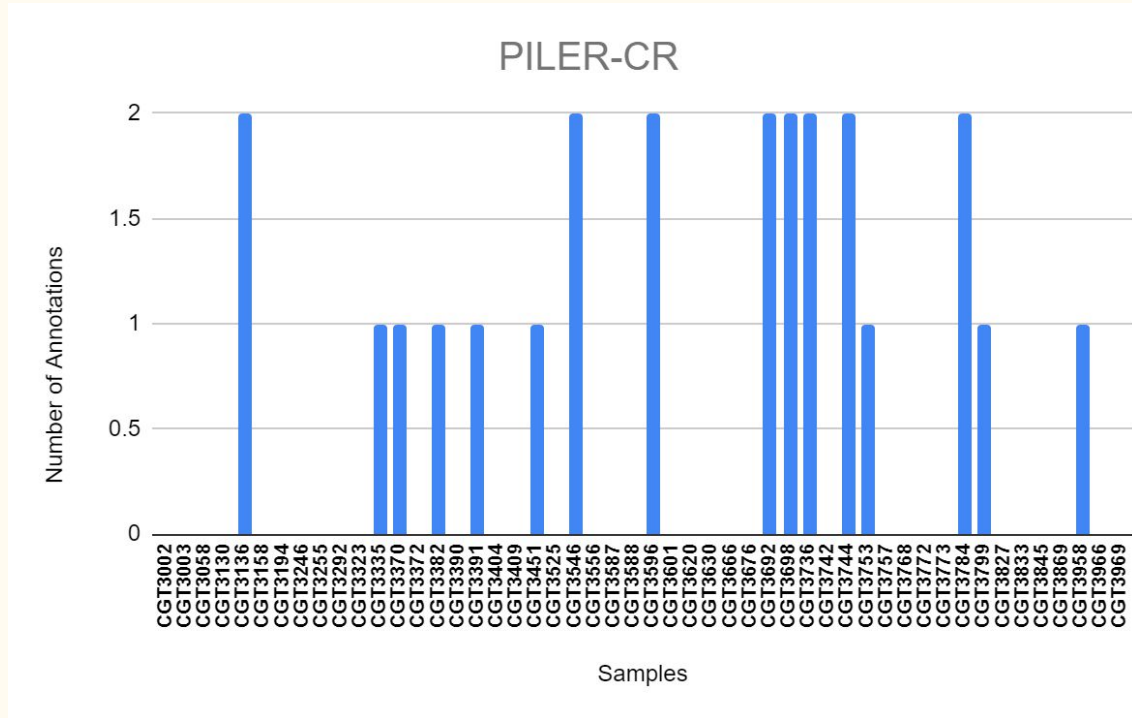
# eggNOG Results

- After mapping the results of eggNog we determined what genes were left unannotated by eggNOG
- We mapped these unannotated genes to the output of InterProScan.
- From this we were able to gain more coverage



# PILER-CR Results

- Identifies CRISPR repeats which play a major role in bacteria's antiviral defense system
- The results shown are from non-coding regions of genomes
- Only one genome had a CRISPR array in coding region of genome
- Command used-  
`./pilerer -in <input_file> -out <output_file>`

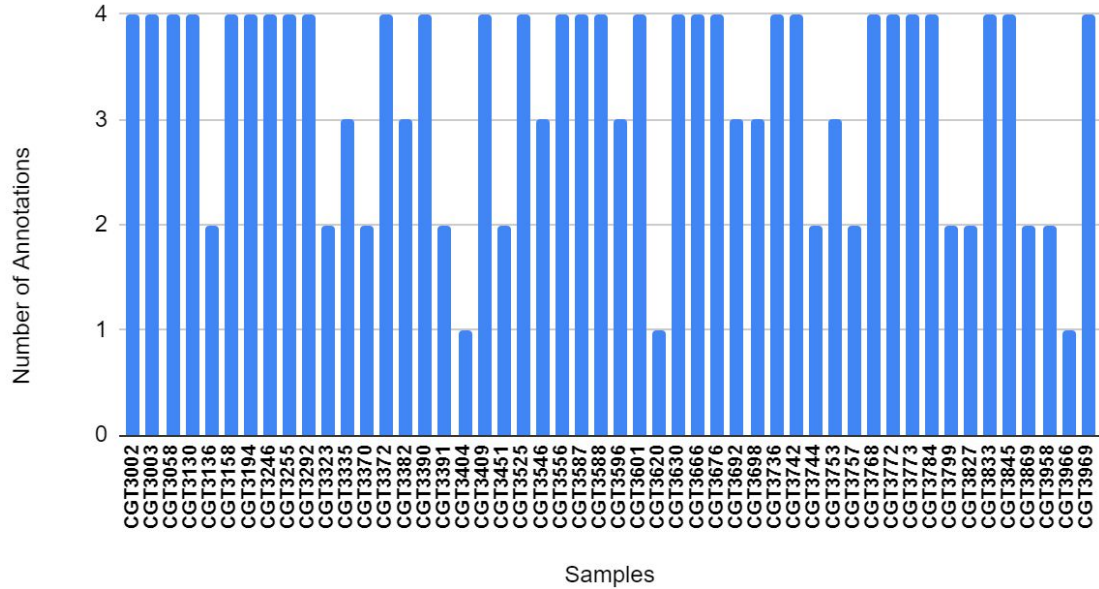




# CARD-RGI Results

- CARD is a rigorously curated collection of characterized, peer-reviewed ARG which is monthly updated
- The results shown are from coding regions of genomes
- No antibiotic resistance genes were present in the non-coding region
- Command used-  
`rgi -i <input_file> -o <output_file>`

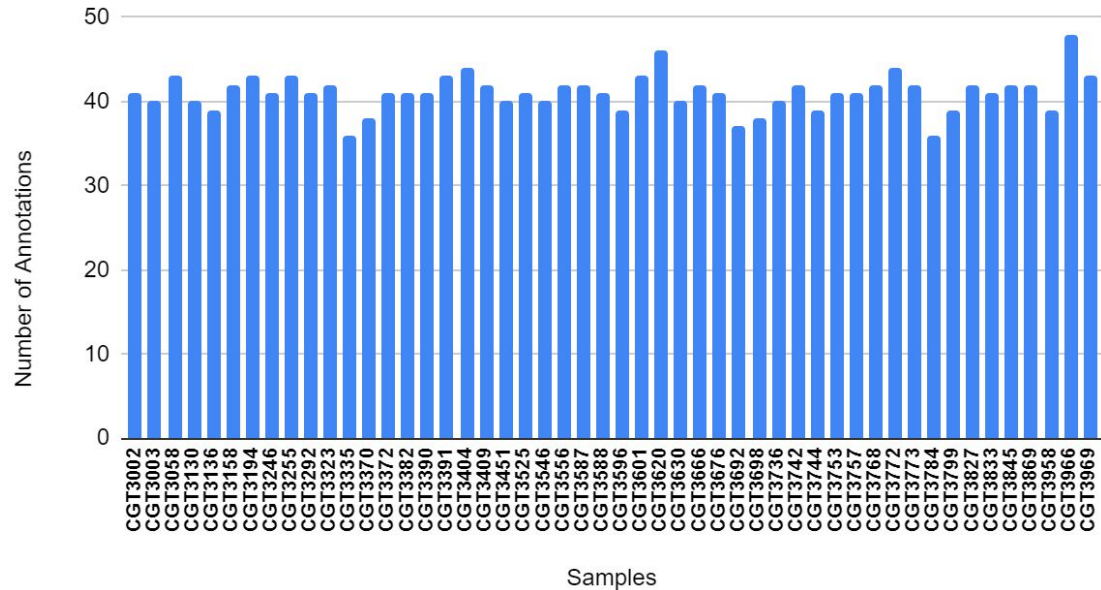
Comprehensive Antibiotic Resistance Database (CARD)



# VFDB Results

- VFDB is an integrated and comprehensive online resource for virulence factors of bacterial pathogens (recently updated in 2019)
- The results shown are from coding regions of genomes
- No virulence genes were present in the non-coding region
- Commands used-

Virulence Factor Database (VFDB)



```
makeblastdb -in <input_db> -parse_seqids -blastdb_version 5 -dbtype nucl -out <name_db>
```

```
blastn -db <name_db> -query <input_file> -out <output_file> -max_hsps 1 -max_target_seqs 1 -num_threads 4 -evalue 1e-5
```

# SignalP

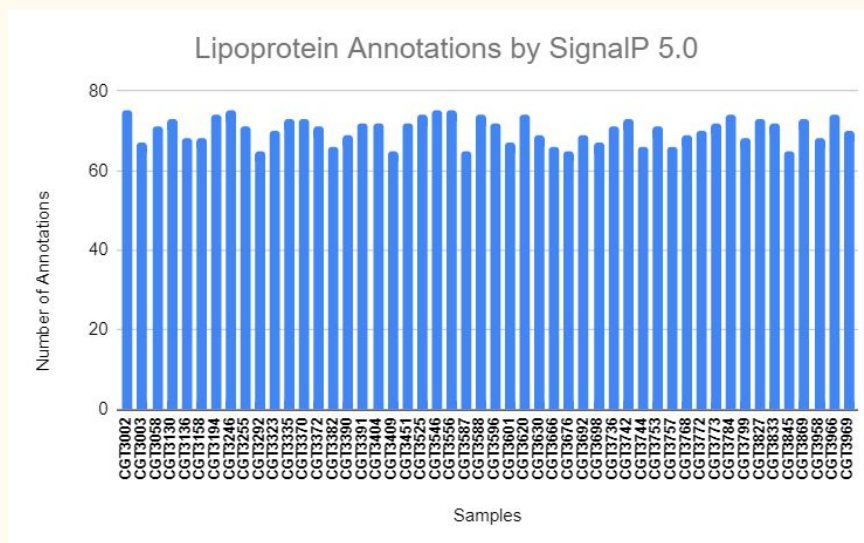
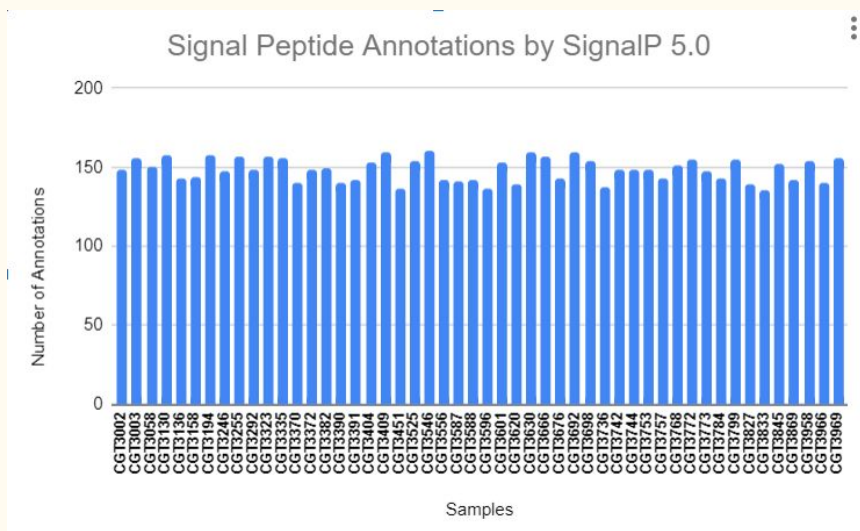
SignalP 5.0 is a deep neural network-based method combined with conditional random field classification and optimized transfer learning for improved SP prediction. [4]

Characterizes between signaling peptides and lipoproteins

Outputs probability of predictions and position of protein in the sequence

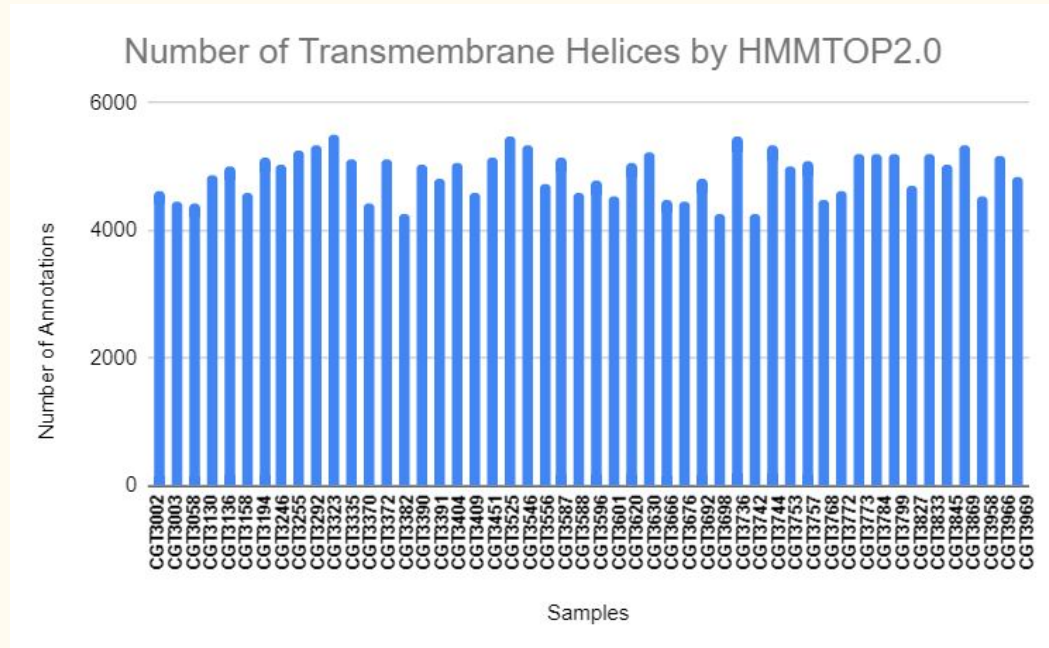
```
# SignalP-5.0 Organism: gram+ Timestamp: 20200327175016
# ID Prediction SP(Sec/SPI) TAT(Tat/SPI) LIPO(Sec/SPII) OTHER CS Position
NODE_1_length_757168_cov_36.966159:121535-121643 SP(Sec/SPI) 0.492041 0.245177 0.205478 0.057304
CS pos: 12-13. AAA-TT. Pr: 0.0605
NODE_1_length_757168_cov_36.966159:125444-126320 SP(Sec/SPI) 0.550027 0.142682 0.284907 0.022383
CS pos: 15-16. ATA-AT. Pr: 0.1476
NODE_1_length_757168_cov_36.966159:127250-128453 SP(Sec/SPI) 0.672480 0.174968 0.134644 0.017909
CS pos: 20-21. AAA-AT. Pr: 0.2309
NODE_1_length_757168_cov_36.966159:140231-141215 LIPO(Sec/SPII) 0.276420 0.108397 0.571096 0.044086
CS pos: 19-20. TAA-CA. Pr: 0.5393
NODE_1_length_757168_cov_36.966159:141538-143329 LIPO(Sec/SPII) 0.297369 0.095134 0.478493 0.129004
CS pos: 18-19. GGG-CA. Pr: 0.2123
```

# SignalP Results



# HMMTOP Results

The HMMTOP transmembrane topology prediction server predicts transmembrane proteins, transmembrane helices, and their start and end positions in the sequence.



```
>NODE_3_length_167022_cov_23.929249:158320-158701      HP:  4  28  35  59  64  88  95 119 124 148 155 179 184 208 215 237 242 266 273 297 3
02 321 328 351 356 380
ATGATAAAATCAGGAGAATATACTTGTATAAATGGGAAAGAATATAAAGTGATTTTAAAAGATAAAAATGGAAAAAGTTATATAAATAAGTGATAAAAAAGAGCCTGATTTCCAAAAGTATGCTGACGGTATTATGAAAAAGAAATTGATTAGAACAAATT
```

# PlasmidSeeker

Command Line: perl plasmidseeker.pl -d ./db\_w20 -i <input.fasta> -b <closest species> -o <output>

```
(T3FN-1) [scheng98@biogenome2020 PlasmidSeeker]$ perl plasmidseeker.pl -d ./db_w20 -i ../../USEARCH/All.fasta -b ../../Listeria -o output.txt
Loading database...
Converting sample reads to k-mers...
Finding coverage of bacterial isolate...
Bacteria median coverage is 2
Bacteria median coverage is too low (less than 3). Higher coverage dataset is needed or use flag --ponly at plasmidseeker.pl line 287.
(T3FN-1) [scheng98@biogenome2020 PlasmidSeeker]$ perl plasmidseeker.pl -d ./db_w20 -i ../../USEARCH/All.fasta -b ../../Listeria -o output.txt --ponly
Loading database...
Converting sample reads to k-mers...
Plasmids done: 8512 of 8514
Clustering results...
Nothing found...
Done!
(T3FN-1) [scheng98@biogenome2020 PlasmidSeeker]$ perl plasmidseeker.pl -d ./db_w20 -i ../../USEARCH/AllNonCoding.fasta -b ../../Listeria -o output.txt
Loading database...
Converting sample reads to k-mers...
Finding coverage of bacterial isolate...
Bacteria median coverage is 2
Bacteria median coverage is too low (less than 3). Higher coverage dataset is needed or use flag --ponly at plasmidseeker.pl line 287.
(T3FN-1) [scheng98@biogenome2020 PlasmidSeeker]$ perl plasmidseeker.pl -d ./db_w20 -i ../../USEARCH/AllNonCoding.fasta -b ../../Listeria -o output.txt --ponly
Loading database...
Converting sample reads to k-mers...
Plasmids done: 8512 of 8514
Clustering results...
Nothing found...
Done!
```

# Phaster

Required complete genome sequence to run online

# IslandViewer

Required gene bank format file or embl format file to search in the database

# References

1. Huerta-Cepas, Jaime, et al. "Fast genome-wide functional annotation through orthology assignment by eggNOG-mapper." *Molecular biology and evolution* 34.8 (2017): 2115-2122.
2. Edgar, Robert C. "Search and clustering orders of magnitude faster than BLAST." *Bioinformatics* 26.19 (2010): 2460-2461.
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6. Barrangou R. The roles of CRISPR-Cas systems in adaptive immunity and beyond. *Curr Opin Immunol.* 2015;32:36–41. doi:10.1016/j.coi.2014.12.008
7. Zhang, Jiayu, et al. "The CRISPR-Cas9 system: a promising tool for discovering potential approaches to overcome drug resistance in cancer." *RSC advances* 8.58 (2018): 33464-33472.
8. Edgar, Robert C. "PILER-CR: fast and accurate identification of CRISPR repeats." *BMC bioinformatics* 8.1 (2007): 18.
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